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(54) Title: ISOLATION AND CHARACTERIZATION OF A *N. CRASSA* SILENCING GENE AND USES THEREOF

(57) Abstract: An isolated nucleic acid molecule encoding for a protein characterized in that it has a silencing activity and comprises a domain responsible for dsRNA interference is disclosed; furthermore expression vectors suitable for the expression of said sequence in bacteria, plants, animals and fungi are disclosed; the invention refers also to organisms transformed by such vectors.

ISOLATION AND CHARACTERIZATION OF A *N. CRASSA* SILENCING GENE
AND USES THEREOF

5 The present invention relates to the isolation and characterization of a *Neurospora crassa* gene encoding for an essential activity in the co-suppression process and to uses and applications thereof in vegetal, animal and fungine fields.

10 The production of transgenic organisms is of large utility both in basic and applied biological research. The transgenic DNA is usually integrated in the genome and transferred as a Mendelian character. However, in various instances, the transgene introduction induces
15 gene silencing phenomena (Flavell, R.B. 1994), i.e. the repression of the expression of the transgene itself and/or of one or more endogenous homologous genes.

 The gene silencing (suppression of gene expression) can act at two levels: transcriptional (trans-
20 inactivation) where transgenes contain sequences homologous to the silenced gene promoter (Vaucheret, 1993); and post-transcriptional (co-suppression) which requires homologies between coding regions (Flavell, 1994; Stam et al., 1997; Baulcombe, 1996).

25 Generally the silencing induced by a transgene requires an almost complete sequence homology (from 70% to 100%) between transgene and silenced gene sequences (Elkind, 1990).

 In the *Neurospora crassa* filamentous fungus, during
30 the vegetative phase, the presence of transgenes induces a post-transcriptional gene silencing phenomenon, named "quelling" (Cogoni et al., 1996).

By using the *al-1* gene (albino 1) (Schmidhauser et al., 1990) as silencing visual marker, many features of the phenomenon have been discovered (Cogoni et al., 1996). Particularly the *al-1* gene "quelling" in *Neurospora* is characterized in that: 1) the gene silencing is reversible further to the loss of transgene copies; 2) the reduction of mRNA basal level results from a post-transcriptional effect; 3) transgenes containing at least a region of 132 base pairs which is identical to the region encoding for the target gene are sufficient to induce the "quelling"; 4) the duplication of promoter sequences is ineffective to induce the silencing; 5) the "quelling" exhibits a dominant behavior in heterocarions containing both transgenic and untransformed nuclei, indicating the involvement of a trans-acting diffusible molecule among the nuclei; 6) the expression of an aberrant RNA transcribed by the transgenic locus is strictly correlated to silencing, suggesting that the "quelling" can be induced and/or mediated by a transgenic RNA molecule.

Therefore homologies between *Neurospora* silencing and plant co-suppression can be pointed out. The gene silencing in *Neurospora* is reversible, as result of transgenic copies instability during mitotic phase; in plants also the co-suppression reversion is associated with the reduction of transgene copy number, resulting from intra-chromosomal recombination during mitosis or meiosis (Mittelstein Scheid et al., 1994; Stam et al., 1997). Thus both in plants and in *Neurospora* the transgene presence is required to maintain the silencing. As in *Neurospora*, a decrease of the mRNA basal level of the silenced gene results from a post-transcriptional

mechanism (Dehio and Schell 1994; van Blokand et al., 1994; de Carvalho et al., 1995). Furthermore to induce the "quelling", transgenes must contain a portion of the silencing target gene coding sequence, being the promoter region ineffective. In plants coding regions with no promoter sequences can induce silencing (van Blokand et al., 1994) and, as in the "quelling", promoters or functionally active gene products are not required for the co-suppression.

One of the similarities between "quelling" and co-suppression in plants is that both mechanisms are mediated by diffusion factors. In *Neurospora* eterokaryotic strains, nuclei wherein the *albino-1* gene is silenced are able to induce the *al-1* gene silencing of the other not transformed nuclei, all sharing the same cytoplasmic environment (Cogoni et al., 1996). In plants the presence of a diffusion factor results from the fact that the co-suppression is effective in inhibiting the replication of Tobacco Etch Virus (TEV), a RNA virus with an exclusively cytoplasmic cycle. The occurrence of highly diffusible factors, which are effective to mediate the co-suppression, has been demonstrated using the grafting technique in tobacco (Palaqui et al., 1997), showing that silenced tobacco plants are able to transfer the silencing to non-silenced plants through grafting.

The fact that "quelling" and co-suppression share all these features suggests that mechanisms involved in post-transcriptional gene silencing in plants and in fungi can be evolved by an ancestral common mechanism.

Recently gene inactivation phenomena resulting from transgene introduction have been disclosed in animals. In *Drosophila melanogaster* the location of a transgene close

to heterochromatic centers results in a variegate expression (Wallrath and Elgin, 1995; Pirrotta, V., 1997). Similar expression profiles have been observed when the reference transgene is within tandem arrayed transposons, indicating that tandem repeats are effective to induce the chromatin condensation. (Dorer and Henikoff, 1994). Again in *Drosophila* Pal-Bhadra et al. (1997) have observed that the transgene introduction can lead to gene inactivation phenomena, similar to the co-suppression.

Gene silencing phenomena resulting from transe gene sequence repeats have been disclosed recently in mammals.

Garrick et al. (1998) produced mouse transgenic lines wherein 100 transgenic copies are present in a unique locus and are repeats-arrayed in direct tandem. The transgene expression has been disclosed to be inversely proportional to the number of occurring copies, indicating that silencing phenomena dependent on repeat copies are present also in mammals.

It has been recently found that double stranded RNA molecules can induce a sequence-specific silencing in several organisms (Fire A., 1999). The mechanism known as dsRNAi (double stranded RNA interference) acts at a post-transcriptional level by inducing sequence-specific degradation of homologous mRNAs (Montgomery, Xu and Fire, 1998). Under this aspect, dsRNAi and quelling in *Neurospora* are similar mechanisms, both of them acting at a post-transcriptional level. In addition, both RNA-induced silencing and DNA-induced silencing can be transmitted from cell to cell.

Therefore the identification of *Neurospora* genes which are involved in the silencing is the first step to modulate the same process in plants, animals and fungi. The silencing modulation is of great relevance when transgenic organisms able to express the desired phenotype are produced.

The authors of the present invention have already isolated *Neurospora crassa* strains mutated at essential functions for gene silencing (Cogoni and Macino, 1997); 15 independent isolated mutants define three complementation groups, thus identifying the *qde-1*, *qde-2* and *qde-3* genes (*qde* stands for "quelling"-deficient), whose products are essential to the silencing machinery. *qde* genes are essential to the *Neurospora* silencing, as suggested by the fact that silencing of three independent genes (*al-1*, *al-2* and *qa-2*) is impaired by *qde* mutations (Cogoni and Macino, 1997).

The authors of the present invention have already identified *qde-3* gene (PCT WO 00/327885) and *qde-1* gene (PCT WO 00/50581).

The authors of the invention have identified and cloned now one out of *Neurospora qde* genes, the *qde-2* gene, thus identifying one of required factors for silencing. By considering the similarity between "quelling" and co-suppression, genes orthologous to the isolated gene are involved in co-suppression and more generally in gene silencing in other organisms, like plants, fungi and animals.

The present invention can be applied with reference to two general scopes: 1) silencing potentiation as a tool for inactivating more effectively and durably a

desired gene, and 2) silencing suppression to obtain a better expression of the introduced transgenes.

The isolated *qde-2* gene can be introduced alone or with *qde-1* and/or *qde-3* genes in plants, animals or fungi, in order to inactivate the expression of selected genes. The aim is to activate a sequence-specific silencing mechanism both in deficient organisms and in organisms wherein the same is not very efficient. The gene silencing can be induced also by introducing specific double stranded DNA or RNA sequences, homologous to the gene to be inactivated.

As to the silencing potentiation, the over-expression of one or more genes controlling the phenomenon can lead to higher efficiency and/or stability thereof. Therefore the introduction of *qde-2* gene or of homologous genes thereof in organisms can constitute a tool to repress more effectively gene functions. Particularly this approach is specially useful in plants wherein the co-suppression is usually used for the "knock-out" of gene functions. In plants again the gene silencing potentiation can be used to obtain lines resistant to pathogen virus, by introducing transgenes encoding for viral sequences, in order to achieve the expression inhibition of the virus itself (Flavell et al., 1994).

Analogous applications are suitable for animals, wherein some indications suggest that silencing can inhibit the suitable expression of introduced transgenes (Garrick et al., 1998).

On the contrary, there are instances wherein it is desirable not to have or to reduce the gene silencing, i.e. where a transgene is to be over-expressed. It is

known that the co-suppression is strictly correlated both with the presence of an high copy number of the transgene, and with a transgene high expression. This correlation can hamper the production of transgenic organisms which express a transgene at high levels, because more high is the expression and/or the copy number, more probable is to evoke silencing responses. As above mentioned, analogous mechanisms of gene inactivation, dependent on a high copy number, have been disclosed in animals. In these circumstances plant or animal lines, totally or partially ineffective for silencing, constitute an ideal recipient wherein the desired gene can be over-expressed. The invention can be applied within this scope using different approaches:

15 A) Identification and production of mutant lines in genes homologous to *qde-2* gene, in plants, animals and fungi.

 The identification of *Neurospora qde-2* gene, essential for silencing mechanism, can allow the isolation of mutant lines in other organisms, mutated in genes homologous to *qde-2*. For example by means of amplifications using degenerated primers, designed from the most conserved regions of *qde-2* gene, mutant lines in homologous genes can be identified, by analysis of insertion mutant gene banks, already available for many plant species. Both in fungi and animals such mutants can be obtained, following the identification of the homologous gene, by means of "gene disruption" techniques using homologous recombination.

30 B) Reduction of *qde-2* gene expression

 Other strategies for the production of silencing-deficient lines comprise the use of *Neurospora qde-2* gene

or homologous genes thereof. *qde-2* or homologous genes can be introduced into suitable expression vectors to express them in an anti-sense orientation in order to inhibit the expression of resident endogenous genes. Alternatively portions of *qde-2* or of homologous genes can be over-expressed, in order to obtain a negative dominant effect and thus blocking the function of *qde-2* endogenous genes.

The authors of the present invention have cloned and characterised the *Neurospora crassa qde-2* gene. The sequence analysis of the *qde-2* gene detected a region having a significant homology with the sequence of a *C. elegans* gene, *rde-1*, involved in the dsRNA mediated interference (Tabara et al., 1999).

The authors of the invention for the first time have demonstrated that the transgene induced post-transcriptional gene silencing and the dsRNA interference share common genetic mechanisms. This supports the hypothesis that the sequence specific gene silencing phenomena evolved from an ancestral mechanism aimed to protect the genome against transposons. Furthermore, the results of the authors suggest that dsRNA molecules are involved in the post-transcriptional gene silencing in fungi. dsRNA molecules could be produced directly from integrated transgenes as a result of the presence of inverted repeats or as an out come of transcription from convergent inverted promoters. Alternatively, single stranded aberrant RNA may be used as a template by an RNA-dependent RNA polymerase (such as QDE-1 protein) able to produce dsRNAs.

Within the scope of the invention the term homology is intended as similarity, i.e. number of identical

residues + number of conserved residues with respect to the total residues of the considered sequence.

Therefore it is an object of the present invention an isolated nucleic acid molecule encoding for a protein
5 characterized in having a silencing activity and in comprising a domain responsible for dsRNA interference, wherein the domain is at least 25% homologous with the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). Preferably the domain is at
10 least 30% homologous with the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). More preferably the domain is at least 38% homologous with the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). Most preferably the
15 domain comprises the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). According to a particular embodiment the isolated nucleic acid molecule encodes for a protein having the amino acid sequence of fig. 1 (SEQ ID No. 2) or functional portions
20 thereof. Even more preferably the isolated nucleic acid molecule has the sequence of fig. 1 (SEQ ID No. 1) or its complementary sequence.

A further object of the invention is an expression vector comprising, under the control of a promoter which
25 directs the expression in bacteria, the isolated nucleic acid molecule of the invention. Those skilled in the art will appreciate that any plasmid suitable for a correct and effective expression of the protein of the expression in bacteria can be used and it is within the scope of the
30 invention.

A further object of the invention is an expression vector comprising, under the control of a promoter which

directs the expression in plants or in specific plant organs, the isolated nucleic acid molecule of the invention, both in a sense and anti-sense orientation. Those skilled in the art will appreciate that any plasmid
5 suitable for a correct and effective expression of the protein of the invention in plants or in specific plant organs can be used and it is within the scope of the invention.

A further object of the invention is an expression
10 vector comprising, under the control of a promoter which directs the expression in fungi, the isolated nucleic acid molecule of the invention, both in a sense and anti-sense orientation. Those skilled in the art will appreciate that any plasmid suitable for a correct and
15 effective expression of the inventive protein in fungi can be used and it is within the scope of the invention.

A further object of the invention is an expression
vector comprising, under the control of a promoter which directs the expression in animals, the isolated nucleic
20 acid molecule of the invention, both in a sense and anti-sense orientation. Those skilled in the art will appreciate that any plasmid suitable for a correct and effective expression of the protein of the invention in animals can be used and it is within the scope of the
25 invention.

A further object of the invention is a prokaryotic organism transformed by using the expression vector active in bacteria of the invention.

A further object of the invention is a plant or a
30 specific plant organ transformed by using the expression vector active in plants of the invention.

A further object of the invention is a plant mutated at the isolated nucleic acid molecule of the invention having a reduced or inhibited silencing activity.

5 A further object of the invention is a fungus transformed with the expression vector of the invention active in fungi.

A further object of the invention is a fungus mutated at the isolated nucleic acid molecule of the invention and having reduced or inhibited silencing activity.

A further object of the invention is a non-human animal transformed with the expression vector of the invention active in animals.

15 A further object of the invention is a non-human animal mutated at the isolated nucleic acid molecule of the invention and having a reduced or inhibited silencing activity.

A further object of the invention refers to a protein characterized in having a silencing activity and in comprising a domain responsible for dsRNA interference, wherein the domain is at least 25% homologous with the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). Preferably the domain is at least 30% homologous with the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). More preferably the domain is at least 38% homologous with the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). Most preferably the domain comprises the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). According to a particular embodiment the isolated nucleic acid molecule encodes for a protein

having the amino acid sequence of fig. 1 (SEQ ID No. 2) or functional portions thereof.

It is within the scope of the present invention the use of the isolated nucleic acid molecule of the invention to modulate gene silencing in plants, animals and fungi.

The present invention now will be described by way of non limiting examples with reference to the following figures:

Figure 1: The isolated nucleic acid molecule of the 5.7 Kb fragment containing the *qde-2* gene and flanking sequences (SEQ ID No.1). The amino acid sequence (SEQ ID No. 2) is shown above the nucleotide sequence.

Figure 2: It is schematically represented the pMXY2 plasmid insertion site, in the 80 mutant, used for insertional mutagenesis and consequent polymorphism of the restriction fragments by mean of DNA southern blot of a WT strain and of 80 and 820 mutant strains by using the entire restored flanking region as probe. The 820 mutant has a complete deletion of the *qde-2* gene.

Figure 3: Multiple alignment, at the conserved region, among *qde-2* and other proteins belonging to ago-elf2C family: *A. thaliana ago-1*; rabbit *elf2C*; *C. elegans rde-1*. Identical amino acids are shown in bold.

MATERIALS AND METHODS

E. coli strains

E. coli strain HB101 (F^- , *hsdS20*(rb^- , mb^-), *supE44*, *recA13*, *ara14*, *proA2*, *rspL20*(str^r), *xyl-5*) was used for cloning.

Neurospora crassa strains and growing conditions

Neurospora crassa following strains, supplied by Fungal Genetic Stock Center (FGSC, Dpt. Of Microbiology,

University of Kansas Medical Ctr. Kansas City, KA) were used:

- Wild type (FGSC 987);
- *qa-2/aro9* (FGSC 3957A), (FGSC 3958a).

5 The 6XW strain (Cogoni et al., 1996) was obtained upon transformation of the FGCS 3958a strain with pX16 plasmid (Cogoni et al., 1996). This plasmid contains the *qa-2* gene used as selective marker and the *al-1* coding sequence.

10 The mutant strains M7, M20 (*qde-1*); M10, M11 (*qde-2*); M17, M18 (*qde-3*) are described in Cogoni and Macino, 1997.

 The *qde* mutants were obtained by UV mutagenesis. As recipient the transforming strain (6xw) silenced at the *albino-1* gene was used. *qde* mutants were selected for their ability to recover a wild type unsilenced phenotype and then classified in three different complementation groups. By analyzing the *al-2* gene quelling frequency all of *qde* used mutants are defective for the general silencing mechanism.

15

20

 Complementation assays with not forced heterocaryons were carried out according to Davis and DeSerres, 1970.

Plasmids and libraries

25 The plasmid pMXY2, disclosed in Campbell et al. 1994, used for insertional mutagenesis was obtained from Fungal Genetic Stock Center (FGSC, Dpt. Of Microbiology, University of Kansas Medical Ctr. Kansas City, KA). The plasmid contains the *Bml* gene (allele responsible of the benilate drug resistance), that was used as selective marker after transformation. The genomic DNA containing

30

the *qde-2* gene was isolated from a *N. Crassa* gene library in cosmids. (Cabibbo et al., 1991).

N. crassa transformation

Spheroplasts were prepared according to the Akins and Lambowitz (1985) protocol.

Southern Blot Analysis

Chromosomal DNA was prepared as disclosed by Ireland et al., 1993. 5 µg of genomic DNA were digested and blotted as reported in Maniatis et al.

DNA probes were: a) as to the *al-1* gene the probe is represented by a XbaI-ClaI restriction fragment of pX16 (Cogoni et al., 1996); b) as to the *BmI* gene the probe is represented by the 2.6Kb SalI fragment of pMX2.

Northern Blot Analysis

N. crassa total RNA was extracted according to the protocol described by Cogoni et al., 1996. The mycelium was grown for two days at 30°C, then powdered in liquid nitrogen before RNA extraction. For Northern analysis 10 µg of RNA were formaldehyde denatured, electrophoresed on a 1% agarose, 7% formaldehyde gel, and blotted over Hybond N (Amersham) membranes. Hybridization was carried out in 50% formamide in the presence of ³²P labeled DNA probe 1.5x10⁶ cpm/ml.

RESULTS

Isolation of silencing mutant by insertional mutagenesis

Previously a *Neurospora* strain (6XW) wherein the *albino-1* resident gene was steadily silenced was used for UV mutagenesis that brought to the isolation of *qde* ("quelling" deficient) mutants in *N. crassa* induced gene silencing (Cogoni and Mancino 1997).

The 6XW strain shows an albino phenotype due to the lack of carotenoid biosynthesis, as results by the

silencing of the albino 1 gene expression (Schmidhauser et al., 1990). A mutation interfering with the silencing machinery is easily detectable by producing a wild type phenotype (bright orange) of the carotenoid biosynthesis.

5 By means of complementation assays it was possible to establish that *qde* mutants belong to three complementation groups, indicating the presence of three genetic loci involved in the *Neurospora* silencing mechanism. In order to isolate the *qde* genes an

10 insertional mutagenesis was carried out with the 6XW strain, previously used for UV mutagenesis. The insertional mutagenesis was carried out by transforming the 6XW strain with a plasmid, taking advantage of the fact that, after the transformation, plasmids are

15 randomly inserted in the *Neurospora crassa* genome. The mutagenesis was carried out transforming the 6XW silenced strain with pMXY2 (see Materials and Methods) which contains the benilate resistance as selective marker. Transformed strains able to grow in the presence of

20 benilate containing medium and showing a wild type phenotype for the carotenoid biosynthesis were selected. Out of 50.000 isolated independent transformed strains, a benilate resistant strain (80) was isolated, which showed the bright orange phenotype expected for a *qde* gene

25 mutation. In order to verify that the silencing release was effectively due to a *qde* gene mutation and not to the loss of *al-1* transgene copies, the genomic DNA of the strain 80 was extracted and digested with *Sma*I and *Hind*III restriction enzymes. After blotting, DNA was

30 hybridized with a probe corresponding to the coding sequence of *al-1*. The *Sma*I site is present only once in the *al-1* transgene containing plasmid and the digestion

by using said enzyme produces a 5.5Kb fragment corresponding to tandem arrayed *al-1* transgenes, while a 3.1Kb fragment is expected from the resident *al-1* locus. The number of *al-1* transgenic copies present in the 80 strain is comparable to that present in the silenced 6XW strain.

The strain 80 is mutated in *qde-2* gene

The strain 80 was assayed in a heterokaryon assay with a wild type strain and with M7, M20 (*qde-1*) M10, M11 (*qde-2*), M17, M18 (*qde-3*) mutants and with a wild strain (Cogoni and Macino, 1997). As shown in Table 1 the *al-1* gene silencing is restored producing an albino phenotype in all of heterokaryons but M10 and M11. This behavior is consistent with the presence of a *qde-2* gene recessive mutation in the strain 80.

Table 1

Reciprocal heterokaryons among the mutant 80 and previously characterized *qde* mutants.

	80	M7	M20	M10	M11	M17	M18
80	WT	AL	AL	WT	WT	AL	AL
M7		WT	WT	AL	AL	AL	AL
M20			WT	AL	AL	AL	AL
M10				WT	WT	AL	AL
M11					WT	AL	AL
M17						WT	WT
M18							WT

WT = heterokaryon with a wild type phenotype for carotenoid accumulation;

AL = heterokaryon with an albino phenotype wherein the *al-1* gene silencing is restored.

Recovery of sequences flanking the pMX2 plasmid integration site

In order to recover sequences flanking the integration site or sites the following methodology was carried out. The genomic DNA of strain 80 was digested with Aat II enzyme. Subsequently the genomic DNA was ligated and the product used to transform *E. coli* cells that was screened in an ampicillin-containing medium. PQc1 plasmid was recovered and a DNA fragment containing sequences flanking the integration site was isolated from it by using Aat II and Cla I enzymes.

10 Isolation of genomic clones, their subcloning and complementation of the *qde-2* mutant

The fragment from pQc1 plasmid was used to probe a *Neurospora crassa* genomic library in cosmids. Three cosmids 6G10, 20C1 and 23F2 containing about 35 Kb genomic DNA inserts, were isolated. Such cosmids were used in transformation experiments of M11 and 80 mutants. All of cosmids are able to restore the *al-1* gene silencing in the two mutants, determining the appearance of an albino phenotype. The 20C1 cosmid was used to subclone a 5.7 Kb BamHI-BamHI fragment. This subclone was used for transformation experiments and resulted to be able to complement the *qde-2* phenotype, indicating that a *qde-2* functional gene is present in this plasmid.

20 Isolation and sequence of the *qde-2* cDNA

25 The sequence of BamHI-BamHI region allowed to deduce the amino acid sequence of the QDE-2 protein. The *qde-2* gene encodes for a 938 aa. putative protein (104 KDa). The genomic clone does not contain any introns since the reading frame does not contain any interruptions and intron acceptor and donor sequences were not identified (Fig. 1, Seq. ID No 1, 2).

30

The qde-2 gene comprises an homologous domain with encoding genes for proteins that are responsible for dsRNA interference

5 The 938 aa sequence (SEQ ID No. 2) was used to search in database of amino acid sequences, by using the BLASTP algorithm. As showed in fig. 3, the search identified significant homologies with argonaute-1 gene [with expected values (E value) of 2e-57] of *A. Thaliana* (mutants of this gene show developmental anomalies); rde-10 1 gene [with expected values (E value) of 1e-23] of *C. elegans*, involved in gene silencing phenomena induced by double stranded RNA; elf2C gene [with expected values (E value) of 5e-60] of rabbit isolated as an element belonging to transcription beginning complex.

15 Plant expression vector

 The qde-2 gene was inserted, in a sense orientation, into a vector containing a plant expression "cassette", including the 35S promoter and the PI-II "terminator" sequences. The vector also includes the20 *Streptomyces hygroscopicus* bar gene, which confers the phosphinotricine herbicide resistance to transformed plants. In an analogous vector to the above mentioned one, qde-2 was inserted in an anti-sense orientation with respect to the 35S promoter.

25 The obtained vectors can be utilized to over-express the qde-2 gene in plants, or to repress the gene expression of resident genes, which are homologous to qde-2.

Fungus expression vector

30 The qde-2 gene was inserted in a vector containing a fungal specific expression "cassette", comprising the *A. nidulans trpC* gene promoter and terminator; both in a

sense and an anti-sense orientation. In addition the vector contains the bacterial *hph* gene, which confers the hygromicine drug resistance. The sense plasmid can be used to over express the *qde-2* gene, whereas the anti-sense plasmid is used to repress the expression of *qde-2* homologous genes in various fungine species.

Mammalian expression vector

The *qde-2* gene was inserted in a vector containing a mammalian specific expression "cassette", including the cytomegalovirus (CMV) promoter and SV40 termination and polyadenylation sequences both in a sense and anti-sense orientation. The vector includes also the neomycine phototransferase gene, as marker for mammalian cell selection. The sense plasmid can be used to over express the *qde-2* gene, whereas the anti-sense plasmid can be used to repress the expression of *qde-2* homologous genes in various mammalian species.

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Claims

1. An isolated nucleic acid molecule encoding for a
5 protein characterized in having a silencing activity and
in comprising a domain responsible for dsRNA
interference, wherein the domain is at least 25%
homologous with the amino acid sequence from aa. 373 to
aa. 910 of SEQ ID No. 2.

10 2. An isolated nucleic acid molecule encoding for a
protein characterized in having a silencing activity and
comprising a domain responsible for dsRNA interference
according to claim 1, wherein the domain is at least 30%
homologous with the amino acid sequence from aa. 373 to
15 aa. 910 of SEQ ID No. 2.

3. An isolated nucleic acid molecule encoding for a
protein characterized in having a silencing activity and
comprising a domain responsible for dsRNA interference
according to claim 2, wherein the domain is at least 38%
20 homologous with the amino acid sequence from aa. 373 to
aa. 910 of SEQ ID No. 2.

4. An isolated nucleic acid molecule encoding for a
protein characterized in having a silencing activity and
comprising a domain responsible for dsRNA interference
25 according to claim 3, wherein the domain is the amino
acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.

5. An isolated nucleic acid molecule encoding for a
protein characterized in having a silencing activity and
comprising a domain responsible for dsRNA interference
30 according to claim 4, wherein said isolated nucleic acid
molecule encodes for a protein having the amino acid
sequence of SEQ ID No. 2, or functional portions thereof.

6. An isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 5, wherein said isolated nucleic acid molecule has the sequence of SEQ ID No. 1 or its complementary sequence.

7. Expression vector comprising, under the control of a promoter that directs the expression in bacteria, the isolated nucleic acid molecule according to any one of claims 1-6.

8. Expression vector comprising, under the control of a promoter that directs the expression in plants or in specific plant organs, the isolated nucleic acid molecule according to any one of claims 1-6, both in a sense and anti-sense orientation.

9. Expression vector comprising, under the control of a promoter that directs the expression in fungi, the isolated nucleic acid molecule according to any one of claims 1-6 both in a sense and anti-sense orientation.

10. Expression vector comprising, under the control of a promoter that directs the expression in animals, the isolated nucleic acid molecule according to any one of claims 1-6 both in a sense and anti-sense orientation.

11. Prokaryotic organism transformed by using the expression vector active in bacteria according to claim 7.

12. Plants or a specific plant organ transformed by using the expression vector active in plants according to claim 8.

13. Plant mutated at the isolated nucleic acid molecule according to any one of claims 1-6 having a reduced or inhibited silencing activity.

14. Fungus transformed by using the expression vector active in fungi according to claim 9.

15. Fungus mutated at the isolated nucleic acid molecule according to any one of claims 1-6 having a reduced or inhibited silencing activity.

16. Non-human animal transformed by using the expression vector active in animals according to claim 10.

17. Non-human animal mutated at the isolated nucleic acid molecule according to any one of claims 1-6 having a reduced or inhibited silencing activity.

18. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference wherein the domain is at least 25% homologous to the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.

19. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 18 wherein the domain is at least 30% homologous to the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.

20. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 19 wherein the domain is at least 38% homologous to the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.

21. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 20 wherein the domain is the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.

22. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 21 comprising the amino acid sequence of SEQ ID No. 2 or functional portions thereof.

23. Use of the isolated nucleic acid molecule according to any one of claims 1-6 to modulate the gene silencing in plants, animals and fungi.

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Frame 1

[illegible]

FIG. 1-1

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F K V H L V T T T K L K V P E N R I F E V T
 TTC AAA GTG CAC CTG GTG ACG ACC ACC AAG CTC AAA GTT CCC GAG AAC CGC ATC TTT GAG GTG ACG
 1323 1332 1341 1350 1359 1368 1377
 W T E P S S N Q N L P S K P Q T W V V K V E
 TGG ACC GAG CCG AGT TCC AAC CAA AAC CTG CCC AGC AAG CCC CAG ACT TGG GTG GTC AAG GTG GAG
 1389 1398 1407 1416 1425 1434 1443
 E S V E T C D F G K V L N E L T T L D P K L
 GAG AGT GTC GAA ACC TGC GAT TTC GGC AAG GTG CTG AAC GAG CTC ACG ACA CTT GAT CCC AAG CTC
 1455 1464 1473 1482 1491 1500 1509
 D G D F P K Y N V E L D A L N T I V T H H A
 GAC GGA GAC TTT CCC AAG TAC AAT GTG GAG CTC GAT GCC CTC AAC ACC ATT GTG ACT CAT CAT GCC
 1521 1530 1539 1548 1557 1566 1575
 R A D D N V A V V G R G R F F A I G D D L I
 CGC GCC GAC GAC AAT GTT GCG GTG GTG GGA AGG GGA AGG TTT TTT GCC ATT GGT GAT GAC CTC ATT
 1587 1596 1605 1614 1623 1632 1641
 E Q V R P H D S P L V I L R G Y F A S V R P
 GAA CAA GTG CCG CCC CAT GAC TCC CCT TTG GTC ATC TTG CGA GGA TAT TTT GCC AGC GTC CCG CCA
 1653 1662 1671 1680 1689 1698 1707
 A T G R L L L N T N I T H G V F R P G V K L
 GCT ACC GGA AGA CTT TTA CTC AAT ACC AAC ATC ACG CAT GGT GTC TTC CGT CCT GGG GTC AAA CTT
 1719 1728 1737 1746 1755 1764 1773
 A Q L F Q E L G L D V M D K C N A W N E V T
 GCA CAG CTG TTT CAG GAA CTT GGA CTT GAC GTA ATG GAC AAA TGC AAT GCC TGG AAC GAA GTA ACC
 1785 1794 1803 1812 1821 1830 1839
 K N Q L N D K M R R V H K V L A K G R V E L
 AAA AAT CAG CTC AAC GAC AAG ATG CGC AGA GTT CAC AAG GTC CTG GCT AAG GGC CGT GTC GAG TTG
 1851 1860 1869 1878 1887 1896 1905
 N A P F L I D G K I V Y K K C Y R T L N G I
 AAT GCC CCA TTC CTT ATT GAT GGA AAG ATT GTT TAT AAA AAA TGT TAC CGC ACG CTC AAT GGC ATT
 1917 1926 1935 1944 1953 1962 1971
 A N R G D E R G K Q K D G K E V R Y P P L F
 GCT AAC CGT GGC GAC GAA AGG GGG AAG CAA AAG GAT GGT AAA GAA GTC CGA TAT CCG CCC TTG TTC
 1983 1992 2001 2010 2019 2028 2037
 G I P G V Q V G G P T S C Q F Y L R A R E T
 GGG ATT CCG GGT GTC CAG GTT GGC GGC CCG ACC TCT TGT CAG TTC TAC TTG CGT GCG CGA GAG ACA
 2049 2058 2067 2076 2085 2094 2103
 K D G A A P P P T P G L P S N A Y I T V A N
 AAG GAT GGC GCT GCC CCT CCT CCG ACT CCC GGC CTG CCG AGC AAC GCG TAC ATC ACG GTA GCG AAC
 2115 2124 2133 2142 2151 2160 2169
 Y Y K Q R Y G I T A N A S L P L V N V G T K
 TAT TAT AAA CAA CGG TAC GGA ATA ACC GCC AAT GCT TCG CTT CCT CTG GTC AAC GTT GGC ACC AAG
 2181 2190 2199 2208 2217 2226 2235
 E K A I Y V L A E F C T L V K G R S V K A K
 GAA AAG GCG ATT TAC GTC TTG GCC GAG TTT TGT ACG CTG GTC AAA GGC CGT TCC GTC AAG GCT AAG
 2247 2256 2265 2274 2283 2292 2301
 L T A N E A D N M I K F A C R A P S L N A Q
 CTG ACA GCC AAC GAG GCG GAC AAC ATG ATT AAG TTT GCT TGC AGA GCT CCT TCG CTG AAC GCT CAG
 2313 2322 2331 2340 2349 2358 2367
 S I V T K G R Q T L G L D K S L T L G K F K
 TCT ATC GTG ACG AAA GGC AGA CAG ACA CTT GGT CTT GAT AAA AGC CTG ACG CTT GGC AAG TTC AAG
 2379 2388 2397 2406 2415 2424 2433
 V S I D K E L I T V V G R E L K P P M L T Y
 GTT TCG ATC GAC AAG GAG CTG ATC ACC GTT GTC GGG CGT GAG CTC AAG CCT CCG ATG CTT ACG TAC
 2445 2454 2463 2472 2481 2490 2499

FIG. 1-2

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S G N K T V E P Q D G G W L M K F V K V A R
 AGC GGT AAC AAG ACG GTA GAG CCG CAG GAC GGC GGG TGG TTG ATG AAG TTT GTC AAG GTC GCC AGA
 2511 2520 2529 2538 2547 2556 2565
 P C R K I E K W T Y L E L K G S K A N E G V
 CCT TGC CGC AAG ATT GAG AAG TGG ACA TAC TTG GAA CTG AAG GGT TCC AAG GCA AAC GAA GGG GTG
 2577 2586 2595 2604 2613 2622 2631
 P Q A M T A F A E F L N R T G I P I N P R F
 CCG CAA GCT ATG ACC GCT TTT GCC GAA TTC TTG AAC AGA ACG GGC ATC CCG ATT AAC CCC AGG TTC
 2643 2652 2661 2670 2679 2688 2697
 S P G M S M S V P G S E K E F F A K V K E L
 TCG CCG GGC ATG AGC ATG TCA GTT CCA GGG AGC GAA AAA GAG TTC TTT GCC AAA GTG AAG GAA CTC
 2709 2718 2727 2736 2745 2754 2763
 M S S H Q F V V V L L P R K D V A I Y N M V
 ATG AGC TCG CAC CAA TTT GTG GTG GTT CTT TTA CCC AGA AAG GAT GTT GCG ATC TAC AAT ATG GTG
 2775 2784 2793 2802 2811 2820 2829
 K R A A D I T F G V H T V C C V A E K F L S
 AAG CGG GCT GCC GAT ATC ACA TTT GGC GTT CAC ACA GTC TGT TGT GTA GCC GAA AAG TTC CTT AGC
 2841 2850 2859 2868 2877 2886 2895
 T K G Q L G Y F A N V G L K V N L K F G G T
 ACT AAG GGG CAG CTG GGG TAT TTT GCC AAC GTC GGC CTC AAG GTC AAC CTC AAG TTT GGC GGC ACC
 2907 2916 2925 2934 2943 2952 2961
 N H N I K T P I P L L A K G K T M V V G Y D
 AAT CAC AAT ATC AAG ACG CCC ATT CCT TTG CTC GCC AAG GGG AAG ACG ATG GTG GTG GGC TAT GAT
 2973 2982 2991 3000 3009 3018 3027
 V T H P T N L A A G Q S P A S A P S I V G L
 GTC ACC CAT CCG ACC AAT CTA GCG GCT GGA CAA TCG CCT GCA TCG GCT CCC AGT ATT GTC GGC CTG
 3039 3048 3057 3066 3075 3084 3093
 V S T I D Q H L G Q W P A M V W N N P H G Q
 GTC TCA ACC ATC GAC CAA CAC CTT GGA CAA TGG CCT GCA ATG GTT TGG AAC AAC CCG CAC GGC CAG
 3105 3114 3123 3132 3141 3150 3159
 E S M T E Q F T D K F K T R L E L W R S N P
 GAG TCC ATG ACG GAA CAG TTT ACG GAC AAG TTC AAG ACG CGT CTG GAA CTA TGG CGC AGC AAT CCC
 3171 3180 3189 3198 3207 3216 3225
 A N N R S L P E N I L I F R D G V S E G Q F
 GCA AAC AAC CGC AGT CTC CCC GAG AAT ATC CTG ATT TTC CGC GAT GGC GTC TCC GAG GGA CAG TTC
 3237 3246 3255 3264 3273 3282 3291
 Q M V I K D E L P L V R A A C K L V Y P A G
 CAG ATG GTC ATC AAG GAC GAG CTA CCC CTG GTT CGC GCC TGC AAG CTG GTG TAT CCA GCT GGC
 3303 3312 3321 3330 3339 3348 3357
 K L P R I T L I V S V K R H Q T R F F P T D
 AAG CTA CCG CGT ATT ACG CTG ATT GTC TCT GTC AAG CGC CAC CAG ACT CGC TTC TTC CCA ACG GAC
 3369 3378 3387 3396 3405 3414 3423
 P K H I H F K S K S P K E G T V V D R G V T
 CCG AAG CAT ATT CAC TTC AAG TCC AAG AGC CCC AAG GAG GGT ACT GTG GTT GAC CGC GGC GTG ACC
 3435 3444 3453 3462 3471 3480 3489
 N V R Y W D F F L Q A H A S L Q G T A R S A
 AAC GTC CGC TAT TGG GAC TTC TTT TTG CAG GCG CAC GCG TCG CTC CAG GGC ACG GCC CGC TCG GCT
 3501 3510 3519 3528 3537 3546 3555
 H Y T V L V D E I F R A D Y G N K A A D T L
 CAC TAC ACA GTT CTG GTG GAT GAG ATT TTC AGG GCC GAC TAT GGA AAC AAG GCG GCC GAC ACG CTG
 3567 3576 3585 3594 3603 3612 3621
 E Q L T H D M C Y L F G R A T K A V S I C P
 GAG CAG CTG ACG CAT GAC ATG TGT TAT CTC TTT GGA CGA GCC ACC AAG GCT GTC AGT ATC TGC CCG

FIG. 1-3

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3633	3642	3651	3660	3669	3678	3687
P A Y Y A D L V C D R A R I H Q K E L F D A						
CCT GCG TAC TAT GCC GAC TTG GTG TGC GAC CGG GCG CGT ATC CAT CAG AAG GAG CTC TTT GAC GCC						
3699	3708	3717	3726	3735	3744	3753
L D E N D S V K T D D F A R W G N S G A V H						
CTC GAT GAA AAC GAT AGC GTT AAG ACC GAT GAT TTC GCA AGA TGG GGT AAC TCC GGG GCT GTT CAT						
3765	3774	3783	3792	3801	3810	3819
P N L R N S M Y Y I						
CCC AAC CTT AGG AAC TCC ATG TAC TAT ATC TAG GCT TGT CAA TTG TGT GCT GGA ATG TAC TGG AGC						
3831	3840	3849	3858	3867	3876	3885
ATA TAA GTG ACG CGA TGG AAG CCT AAT CGT CTC TGA ATA TGG ATC AAA GAC AGC GTT TGC TTT TTC						
3897	3906	3915	3924	3933	3942	3951
GGG GCT TCT AGT TTC TAC AGC GAT TTG TGT GGA TTG TTT CTT GTT CTG TTT CTT GGT TCT TTC TTT						
3963	3972	3981	3990	3999	4008	4017
CTT TTT TTT GTG TCT CTG TCT GCC TTT GTA CTG CAT GCA AAC GTG CAC TCT GAA TGA TGA ACG ACA						
4029	4038	4047	4056	4065	4074	4083
CCA TTT GAC GAT TGG ATA AGA GAT GAC AGA CTG CAG ATA CTA TCA TGC GCA ATG GAA AAC ACG AAC						
4095	4104	4113	4122	4131	4140	4149
AAC CAA GGT TTT TGA TTC CTT CAA TAG CGA AAT ATA GAA AAA GAA ACA AAA AAA AAA ACA ACA ACA						
4161	4170	4179	4188	4197	4206	4215
AAT AAT GGA AGT ATG ATT AAA CAC ATT GAG CGC GAT GAC TGA CTG GTG TTG TGA ATG GCG TGT TGG						
4227	4236	4245	4254	4263	4272	4281
TTT TCT TCT TTC TTG AAA ATT TAG AAC CGT AAA TGT TAT ATC ATG TGA TGT AAT GTA ATA ACA TAT						
4293	4302	4311	4320	4329	4338	4347
TTA TAT CTC GTT GTA TTC TTG TAC ACA CTT TCC AGG ATA ACA TGG TCT GAC ATG GTA TTT CTG ACG						
4359	4368	4377	4386	4395	4404	4413
TAC AAA AAA GAA AAA GAA AAA CAG GAA ACC ATG AAC CCG CGA CAA AGC TGT TCC AGT TGT TAC AAT						
4425	4434	4443	4452	4461	4470	4479
GAT GAT GAT GAT GAT GAC CTA CTA CCT AAG GTA TTC TAT CTT AGC CAA GGT ATT CTC TCG CAT CCT						
4491	4500	4509	4518	4527	4536	4545
ATT CCA TCC TAT CCT AAC CCG AGC CTA ACC CGA GCC TAA ATA CCT AAA CTC CTA AAC TCC TTA ACT						
4557	4566	4575	4584	4593	4602	4611
CCT TAA CTC CTT TCT AAA TGT CTA AAC CCC CAA ACT ATG AGA CGA CCC GAA CCC GAA ACC CTA ATA						
4623	4632	4641	4650	4659	4668	4677
AAA GTA TTT ATA AAC CAT CAT AAA AGA AAA AAA ACC ATC ATA CAT GGA TGA TCA AAA CAA ACA GAA						
4689	4698	4707	4716	4725	4734	4743
ACG GAA ACA ACA CAA CCA GCT ACC CGC TCA AGA CTT TCA TTC GTT AAT TCA TCA CTC ACT CAC TCA						
4755	4764	4773	4782	4791	4800	4809
CTC ACT CAC TCA GCA GCA AAA TAC CGT TTT GTC CTG CTA TTC GTT TGT TGC GCC TTG ATT TCA GGC						
4821	4830	4839	4848	4857	4866	4875
GGG ACA ATG GTG TGA TGT ACG ACG TGG GGG CGG TAG ACT GCG TCT ACT GGT GGC ATC CTT TAC AAT						
4887	4896	4905	4914	4923	4932	4941
TTT TTA GTG TGT CAG TAT GTG ATG TAT TCA ATG CTA TTG AAC TGA GGG GGG CTG ATG GAT AGT GGG						
4953	4962	4971	4980	4989	4998	5007
GAG AGA ACA CCT GAC GGA TAG AGG GAA GGA ACT GGA CGC CTG GGG GGA AGT GAG AGA GGG GGA TGG						
5019	5028	5037	5046	5055	5064	5073
TGG GGA ATA GAT GAA AAG AGA AGA GGA GTG AGA GCA CAA GAA GAA AGA ATG AAT GTT GGT GAC AAA						
5085	5094	5103	5112	5121	5130	5139

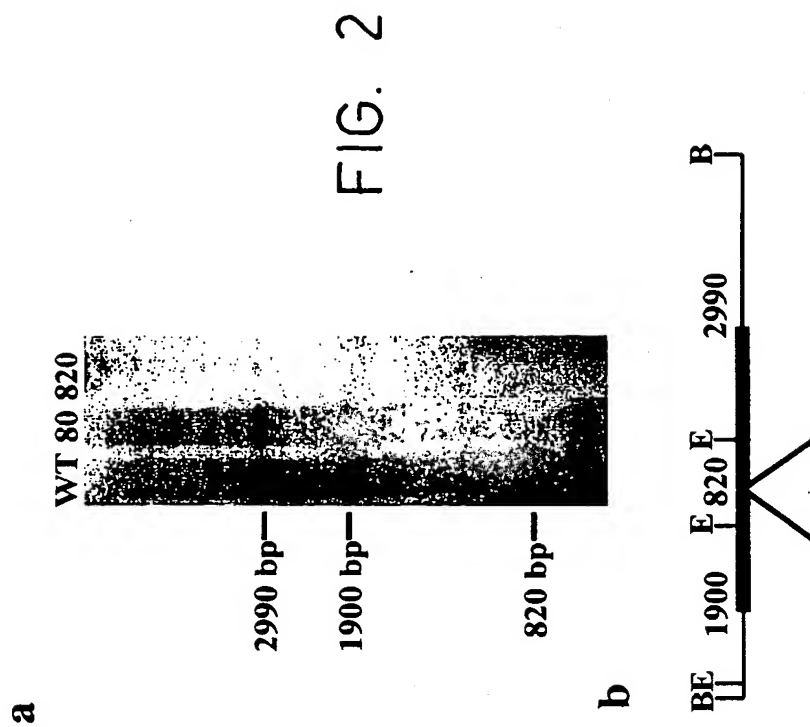
FIG. 1-4

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GTT AAA GAA AAG GAA GGG GGG AAA GAG AAG AGG ACA GGT GTG GTG AGT GAA TTG AGT GAA AGG AAG
 5151 5160 5169 5178 5187 5196 5205
 GGA AAA AAC GGA GAA GGA AAA AAA AAA CAT AAA AAA AAA AAA AAA AAC AGA AAG AAA GAA CTA ACC
 5217 5226 5235 5244 5253 5262 5271
 AAT CAT CCA AAC TCA GCG GAA AGT ACT CAT ACA AAA GGT CGG CTG CCT CAA TCG GAC TCC CCA CAT
 5283 5292 5301 5310 5319 5328 5337
 TCT CTT TCT GGT ACT GAT TCT GCT GCC CCA GAC TTC CAC TTT CAA AGT GGC TAT CAC CCT TAT TGT
 5349 5358 5367 5376 5385 5394 5403
 TGT TAG AGT GAG TAG TAG ACG TAA GTC CTC CCG ATC CGG AGC CAA AAC CCA TCC CTT TCC CAG CTG
 5415 5424 5433 5442 5451 5460 5469
 TAT CCC TCT TCA ATC CAC CAG TAG CAA CAC CCA TCT TGC CAT AGA GCG GAC TAT CCC CTG CCC CTG
 5481 5490 5499 5508 5517 5526 5535
 CCC CTG CCG AGC CAG GAG TAG CAG TCC TAT TCA TAG GCG GAC TCC TCT GCT CGT CTT CCG ACA GGG
 5547 5556 5565 5574 5583 5592 5601
 ACA AAC TAA TTG GTA GGG CAC CCG CAG CAG AGG AGG AGG TAT TTC TGT GAT GAC TGG TTC TGT TTG
 5613 5622 5631 5640 5649 5658 5667
 GGG CAG CTA AGG GCG TGG GTT TCC TTC GTG AGC CGC TGT TGT GAT TGT TGG CGG CGG CGT CCG AGG
 5679 5688 5697 5706 5715 5724 5733
 ATA AGG ATC C
 5745

FIG. 1-5

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AGO-1 LGSRLPAYDGAKSLYTAGFLPFNSKEPRINLLDEYVAGGQRREREFTKVHILVARADLH 300
 eIF2C FGDRKPVTDGKQNLTYAMPLFIGRE-----KVELEVLPCEGKDRIFKVSINWVSCVSLQ 95
 QDE-2 LTTLDPKDDGDFPKVVELDALNTIVTHARADDHNAVVOGRGFFAIGDDIEQVRPHDS 210
 RDE-1 LTYRKKFHLNFSRENPEKDEEANR---SYKFLKQVATQKVRYAPFVNEEHVQFAQNFV 216

AGO-1 HICMFLECKQSDAPQALQVVIDIVRELPTSRYIPVGRSISYSPDIGKKQSLGDLGSSWRG 360
 eIF2C AHDALSGRLPSVDFETIQALDQVVRHLPSMRYITPVGRSFTASEGCSNPLGGGREGVWFG 155
 QDE-2 PIVILRGYFASVRATGRLLINNTHTGVTRPGVKLAQLQELGLDQVMDKCNANWNTKN 270
 RDE-1 YDNNSILRVPSFHDNRFEQSLEVAPRIEAWFGIYIGIKELFDGGEFVLNFAIVDKLYN 276

AGO-1 FYQSIRPTOMGLSLN--IDMSSTAFIEANPIQFVCDLNRDISS---RPSDADRVKIK 415
 eIF2C FHQSVRPSLWKKMLN--IDVSATAFYKAQPIEFVCEVDFKSIIEEQHFTDSORVFT 213
 QDE-2 QLNDKRRVHVKLAKRVELNAPFLIDCKIYKKCYRTINGIANRGDEKQKQCKEYRY 330
 RDE-1 APMSILDYILLIVD--PQSCNDDVKDLKTLKLAGKRTIROAARPRIQOLENLKICA 334

AGO-1 KALREYKVEVHRGNMRRKQIRISGLAVATRELTPVDERN---TQKSVLETHETYGFR 472
 eIF2C KEIKELKVEIHCCGQCRKQRCVNVRRPASHQTFPLQESGQTVECTVAQIKTKRHLV 273
 QDE-2 PPLFETPGVQGGPTSCQFYLRAREKDGAAFPPTPLPSN---AYITVANKYQRYGIT 387
 RDE-1 EVMONEMSRLEERHLEFLDLCEENSLVYKVTGKSDRGNNK---KYDTLFLKIYENKFK 391

AGO-1 IQHTOLPCLOVONSNAIPNLYPMEVCKMVC-DRYSKLENEROITALLVFCORPIL-REK 530
 eIF2C LRYPHLEPCLOVOCQCHTYVPLEVGNVAG-ORCIKLETONOTSTMTATARSAPL-RQE 331
 QDE-2 AN-ASLPLVNVCTKEKAIVYLAETCTAVK-ESVKARITANEADNMKFPACRPSLNAGS 445
 RDE-1 IEFPHLEPLVVKSGAREYAVDMHLEVHEKFORVKNIDLVMDQKFLKATKCHETKEN 451

AGO-1 DILQTVQNDYAKDN-YAQENCKISTSLASVEARHIPPVKKHESGREGTCLQVSCM 589
 eIF2C EISKLMRSASVNTDP-YVREFQVAKDETDVAGVLPQPSSELGGRNK-ALATEVQCVV 389
 QDE-2 IYTKGRQTLGLDKSL-TLGNKYSIDKLEITVGRLEHPP-ITISGKNT--VE-PQGGG 501
 RDE-1 TLQMLKELDFSSSEELNFVERECJCSKLOIECPGVKES-LVNSVNEQIKMTFVIRQ 511

AGO-1 NDNKKMIN-GGTVMNIGICINFS--RQVQNLARTCQELQACVYSCALNPEFVLPV 646
 eIF2C DNRNKQFHT-GIEIKVNAIACFAPQROCTVELKSETEQPKISRDACMTQGGPCFKY 448
 QDE-2 LMKFVKVARPCRKEKTYLELK--CSKANEVGPQANTAFELNRTGIPNFRPSFGMS 559
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 RDE-1 VGFVKIQTHTIGVANCHITSEVYKALASLRHEKSKRITYQALKINAKLGGINQ 691

AGO-1 DALSRIP-----LVSDRPIITICADVTHP-----PGEDSSPSIAAVASQWPEITK 788
 eIF2C PQGR--P-----FVFQPVFLGADVTHP-----AGDGKPSIAAVGSAHPN-R 578
 QDE-2 TPIP-----LLAKGHTVVCGDVVHTPLAAGQSPASAPSVGLSTIDQHLG-Q 696
 RDE-1 WSEIAEISPEEKERRKTMPLATVGVTHPT-----SYSGIDYSIAAVASINPGGT-I 745

AGO-1 VAGLVCAQAPDELQDLFKWQDPQGVVTCGMKELIARRSTCH-KELRHIDYDE 847
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 QDE-2 WPAVWNNPQESMITEQFTDKFKTR-----LEWRSNPANNRS-LEENHIDYDE 746
 RDE-1 VRMDVTEECRPGERA VAHGRETD---ILEAKVKEPRLEENNDNRABAHIVYDE 802

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 eIF2C VSEGOYOILLYE-DALTRKACIKLEK-----YQGITETVVQKRHHKLECTDKHER-- 677
 QDE-2 VSEGOYOILLYE-PLYRAACKLVYPAG-----KLEPITIVSVKRHHKLEPTDPKHIEH- 801
 RDE-1 VSDSEMLRVSNDERSLSEVKQFMSERGEDPEKTYTEIVQKRHHKLELRHEDKQPV 862

AGO-1 V-----DRSCILPCTVVDISKCHPTEDSILCHAG 934
 eIF2C V-----GKSCIPACTVVDKTHPTEDSILCHAG 709
 QDE-2 V-----FKSKSPKGTVDRCYTNVRYEDSILCHAS 832
 RDE-1 VNKDLTPAETDVAVAAVKQWEEDKLSKETGVNPSSTVDKLVSKYKEDSILCHAG 922

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 QDE-2 CDARINHQELFDALD-----END-SVKTDDFARWGNSGAVHPNLNRSYYI- 938
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FIG. 3

SEQUENCE LISTING

<110> Università degli Studi di Roma La Sapienza

Cogoni, Carlo

Macino, Giuseppe

Catalanotto, Caterina

Azzalin, Gianluca

<120> Isolation and characterization of a N. crassa silencing
gene and uses thereof

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<151> 2000-01-17

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ctgatatcga catatcacc cacaacatca tcatcatcta ctaccagtaa tcccgcacgc 180

gaggagtagt cgtttcgctc gattactctt ttttttgctt ccggagtgcg acaaagtagc 240

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2/12

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Leu Pro Ser Lys Pro Gln Thr Trp Val Val Lys Val Glu Glu Ser Val	
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